

Effects of elevated CO₂ and irrigation on gas exchange and water relations among two wheat cultivars

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Abstract

Gas exchange and water relations of two *Triticum aestivum* L. cultivars (Yitpi and H45) were compared under two CO₂ levels (ambient aCO₂ ~380 μmol mol⁻¹, elevated eCO₂ ~550 μmol mol⁻¹) and two water treatments (rain-fed, rain-fed plus irrigated). Yitpi, a high tillering cultivar, was compared to H45, a low tillering cultivar, to study intra-specific variability aspects of responses to a rising atmospheric CO₂ concentration. Measurements were made on flag leaves of plants grown in a Free Air Carbon Dioxide Enrichment (FACE) site during anthesis (DC65-DC68). Both cultivars had increased light saturated net assimilation (A_{sat}) and stomatal conductance (g_s) under eCO₂. When plants grown at either aCO₂ or eCO₂ were measured at a common CO₂ concentration (550 μmol mol⁻¹) to study long-term photosynthetic acclimation, g_s remained greater in eCO₂ grown plants, but A_{sat} was similar. Irrigation increased A_{sat} and g_s as well as decreased cell sap osmolality in Yitpi but there was no irrigation effect in H45. It remains to be seen whether such differences translate into yield variations over a whole growing season.

Key Words

Acclimation, photosynthesis, cell sap osmolality, RWC, climate change

Introduction

Current climate change projections predict a rise in global atmospheric CO₂ concentration (Forster et al. 2007). At the same time, probability and frequency of drought events are expected to increase across an increasing amount of land (Mpelasoka et al. 2008), particularly in dry-land areas important to global wheat production.

In plants, rises in global atmospheric CO₂ concentration commonly increase net assimilation rates (A), while underlying processes of photosynthesis (photosynthetic capacity) are often reduced through continued exposure to elevated atmospheric CO₂ (eCO₂), including a small reduction in A . This reduction in photosynthetic capacity is termed acclimation that occurs over a growing season as plants become so-called acclimated to the eCO₂ (Ainsworth and Rogers 2007). Acclimation is caused by morphological, biochemical and molecular adjustments to growth under elevated CO₂. Drought-induced decreases in plant productivity and growth are linked to decreased photosynthetic rates, which is caused by the restriction of CO₂ diffusion through closing stomata to the site of carboxylation (Flexas et al. 2006). eCO₂ may counteract negative effects of drought, because drought induced lower stomatal conductance (g_s) becomes less restrictive to photosynthesis. For example, Seneweera et al. (2001) reported that eCO₂ reduces canopy transpiration and improves soil moisture availability, because plants operated at lower g_s and showed increased osmotic adjustment.

To secure global food supply in a changing climate, it will be necessary to breed cultivars best suited to cope with drought under eCO₂. This will be most effective if research is conducted directly within the

production areas under free air CO₂ enrichment (FACE) (Ainsworth et al. 2008). To deepen our understanding of intra-specific physiological responses to potential climate change factors, we specifically investigated photosynthetic responses and water relations of two *Triticum aestivum* L. cultivars (Yitpi and H45) to the combined effects of two CO₂ levels (ambient aCO₂ ~380 μmol mol⁻¹, elevated eCO₂ ~550 μmol mol⁻¹) and two water treatments (rain-fed, irrigated). Yitpi is considered a high tillering cultivar, whereas H45 is considered a low tillering cultivar, a difference that can influence the use of assimilates ('sink strength') and potentially causes different responses to the experimental conditions. Plants were grown within a major dry-land wheat production area of Australia, at the Australian Grains Free Air CO₂ Enrichment (AGFACE) site in Horsham, Victoria (Mollah et al. 2009). The aim of our work was to identify variation among cultivars in response to experimental conditions to identify traits beneficial in a future high CO₂ world.

Materials and Methods

Plant material and growth conditions

Yitpi and H45 were grown in a randomised complete block design with 4 replications. Elevated CO₂ blocks were fumigated to an elevated target CO₂ of 550 μmol mol⁻¹. Each block was split for irrigation ('rain-fed' and 'irrigated'). Sowing date was 23 June 2009.

Gas exchange measurements

Light saturated gas exchange was measured on flag leaves during anthesis (DC65-DC68) using an open gas exchange system (Li-6400, Li-Cor, Lincoln, NE, USA). Measurements were carried out with an air flow rate of 250 μmol s⁻¹, a block temperature of 20 °C, and at a saturating light intensity of 1800 μmol m⁻² s⁻¹ PPFD. To evaluate gas exchange responses under respective growing conditions (aCO₂ or eCO₂), photosynthesis was recorded at 390 and 550 μmol mol⁻¹ CO₂. To evaluate the acclimation effects of growth CO₂, comparisons were made at a common CO₂ concentration of 550 μmol mol⁻¹ according to Gutierrez et al. 2009.

Plant water status

Relative water content (RWC) of flag leaves was calculated as (FW – DW) / (TW – DW) where FW is the fresh weight of the leaves recorded immediately after harvesting, TW is the turgid weight after rehydrating leaves to full water saturation, and DW is the dry weight after drying leaves at 70 °C for 60 hours. For the determination of cell sap osmolality, rehydrated leaves were cut into small pieces and shock-frozen in liquid nitrogen. Cell sap of thawed samples was expressed by centrifugation and its osmolality (osmols kg⁻¹) was measured with an osmometer (Osmomat 030, Gonotec GmbH, Berlin, Germany).

Statistical analyses

Data were processed and analysed using SigmaPlot 11.0 and PASW Statistics 18 software. A three-way ANOVA was conducted with cultivar, CO₂ and water treatment as fixed factors. Homogeneity of variances was checked with the Levene's test. Distribution of residuals was graphically checked for deviations from normality.

Results and Discussion

Growth at eCO₂ significantly increased light saturated net CO₂ assimilation rates (A_{sat}) by up to 60 % in both cultivars (C effect, Figure 1a), whereas irrigation affected Yitpi rather than H45, with Yitpi grown under irrigated and eCO₂ conditions responding with the greatest A_{sat} rates (I x Cu effect, Figure 1a). Growth at eCO₂ also significantly increased g_s (C effect, Figure 1b), whereas irrigation again affected Yitpi rather than H45, with Yitpi grown under irrigated and eCO₂ conditions showing the greatest g_s (I x Cu effect, Figure 1b).

When plants grown at aCO₂ were measured at the common eCO₂ level of 550 μmol mol⁻¹ CO₂ to evaluate long-term acclimation, A_{sat} between CO₂ treatments was not different (Figure 2a), whereas the cultivar by irrigation interaction (I x Cu effect, Figure 2a) remained significant, with irrigated Yitpi showing the greatest A_{sat}. There was still a CO₂ effect (C effect) and irrigation x cultivar (I x Cu effect) on g_s when aCO₂ grown plants were measured at 550 μmol mol⁻¹, which again was mostly due to the high values obtained for Yitpi grown under eCO₂ and irrigated conditions (Figure 2b). Cell sap osmolality was significantly greater in rain-fed plants (I effect) as well as in Yitpi compared to H45 (Cu effect, both Figure 1c). There was no significant CO₂ or water treatment effect on the RWC of sample plants (data not shown).

Our results showing increases of A_{sat} in response to growth under eCO₂ are in accordance to a large number of publications, even if the extent of such an increase varies considerably. For example, reported increases in A_{sat} range from ~12% (Sicher and Bunce 1997), ~30% (Garcia et al. 1998), to ~50% (Miglietta et al. 1996) with the latter being in the range of what we found in the present study (60%). Species related differences can be excluded as all these studies were performed on wheat at a similar range of CO₂ and under (Mini)FACE conditions. Garcia et al. (1998) suggested that such differences must result from climate pre-history, differences in soil etc.

Acclimation as a process of optimization of photosynthesis in response to eCO₂ would result in lower rates of A_{sat} if plants are measured at a common CO₂. Such acclimation has been reported widely for wheat systems (e.g. Gutierrez et al. 2009). In contrast, our data give no indication of photosynthetic down-regulation under eCO₂ which agrees with Garcia et al. (1998) or Miglietta et al. (1996) who suggested that under FACE conditions the stimulation of photosynthesis by eCO₂ persists without any evidence of acclimation.

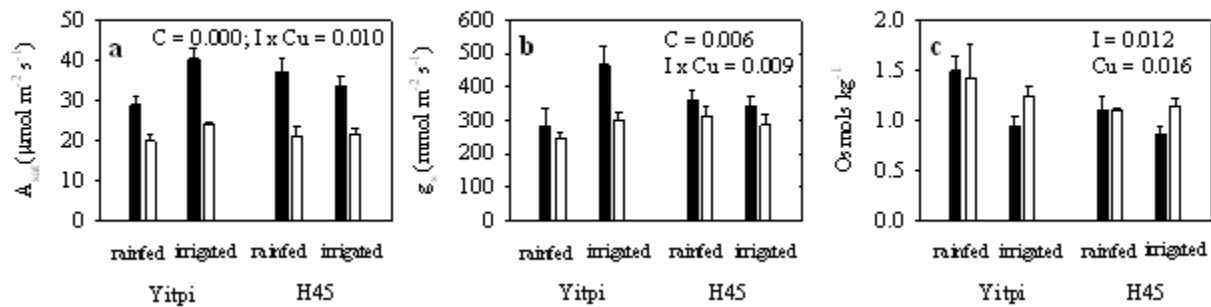


Figure 1. Flag leaf gas exchange (1a and 1b) and water relations (1c) of two *Triticum aestivum* cultivars (Yitpi and H45) grown under two CO₂ levels (closed bars, eCO₂ 550 μmol mol⁻¹, open bars, aCO₂, 390 μmol mol⁻¹) and two water treatments (rain-fed/irrigated). Measurements were taken at growth CO₂ concentrations (390 vs 550 μmol mol⁻¹CO₂). ANOVA effects: I irrigation, C CO₂, Cu cultivar, I x Cu irrigation x cultivar interaction

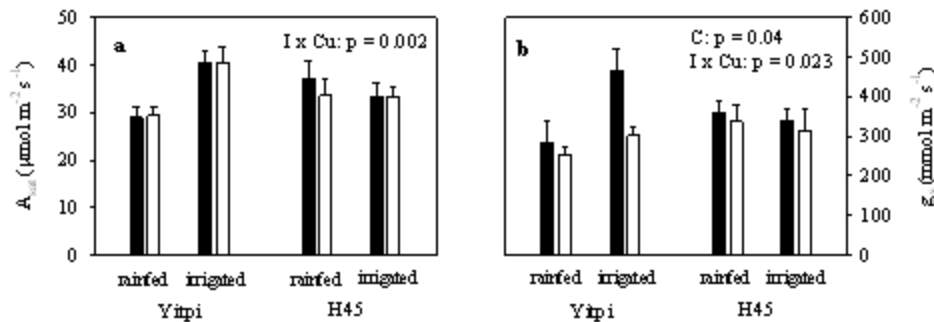


Figure 2. Acclimation responses of Yitpi and H45 to growth under eCO₂. Gas exchange measurements of A_{sat} (2a) and g_s (2b) were made at a common CO₂ concentration of 550 µmol mol⁻¹ CO₂. Closed bars: eCO₂ 550 µmol mol⁻¹; open bars: aCO₂, 390 µmol mol⁻¹. The same experimental conditions as described in Figure 1 apply.

It is overwhelmingly evident from FACE and non-FACE experiments that g_s decreases in eCO₂ (Ainsworth and Rogers 2008), however, there are exceptions due to environmental interactions. For example, Leakey et al. (2006) studied acclimation of g_s under FACE conditions on *Glycine max* by measuring g_s throughout the growing season. They found considerable variation of the CO₂ effects on g_s and they speculated that soil water content becomes less depleted during dry periods under elevated CO₂, thereby potentially minimizing a drought induced decrease in g_s. Similar results were reported by Seneweera et al. (2001). In the present study, we have no direct evidence of greater soil water retention under eCO₂ and there were no differences in RWC of the crop. However, wheat growing under rain-fed conditions showed greater cell sap osmolality than wheat grown with additional irrigation, an indication of osmotic adjustment under dryer conditions (Zivcak et al 2009). Furthermore, there was also a (nearly significant) trend in osmolality towards a CO₂ x irrigation interaction (C x I effect: p = 0.075, data not shown) with eCO₂ grown irrigated plants having lowest values in osmolality possibly indicating greater soil moisture retention within the respective plots.

Conclusion

Overall, we saw substantial stimulation of A_{sat} under eCO₂ and no indication of photosynthetic acclimation. Differences between the investigated cultivars come into play in their responses to different water supply levels, less so in their response to eCO₂. It remains to be seen how such differences reflect on full season growth and yield.

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References

- Ainsworth EA and Rogers A (2007). The response of photosynthesis and stomatal conductance to rising CO₂: mechanisms and environmental interactions. *Plant Cell and Environment* 30, 258-270.
- Ainsworth EA, Beier C, Calfapietra C, Ceulemans R, Durand-Tardif M, Farquhar GD, Godbold DL, Hendrey GR, Hickler T, Kaduk J, Karnosky DF, Kimball BA, K^orner C, Koornneef M, Lafarge T, Leakey ADB, Lewin KF, Long S, Manderscheid R, McNeil DL, Mies TA, Miglietta F, Morgan JA, Nagy J, Norby RJ, Norton RM, Percy KE, Rogers A, Soussana J-F, Stitt M, Weigel H-J and White JW (2008). Next

generation of elevated CO₂ experiments with crops: a critical investment for feeding the future world. *Plant Cell and Environment* 31, 1317-1324.

Flexas J, Bota J, Galmes J, Medrano H and Ribas-Carbo M (2006). Keeping a positive carbon balance under adverse conditions: responses of photosynthesis and respiration to water stress. *Physiologia Plantarum* 127, 343-352.

Forster P, Ramaswamy V, Artaxo P, Bernsten T, Betts R, Fahey DW, Haywood J, Lean J, Lowe DC, Myhre G, Nganga J, Prinn R, Raga G, Schulz M and Van Dorland R (2007). Changes in atmospheric constituents and in radiative forcing. In *Climate Change 2007: The Physical Science Basis. Contribution of working group I to the fourth assessment report of the intergovernmental panel on climate change*. Eds S Solomon, D Qin, M Manning, Z Chen, M Marquis, KB Averyt, M Tignor and HL Miller. Cambridge University Press, Cambridge, United Kingdom and New York.

Garcia RL, Long SP, Wall GW, Osborne CP, Kimball BA, Nie GY, Pinter Jr PJ, Lamorte RL and Wechsung F (1998). Photosynthesis and conductance of spring-wheat leaves: field response to continuous free-air atmospheric CO₂ enrichment. *Plant Cell and Environment* 21, 659-669.

Gutierrez D, Gutierrez E, Perez, P Morcuende R, Verdejo AL, Martinez-Carrasco R (2009). Acclimation to future atmospheric CO₂ levels increases photochemical efficiency and mitigates photochemistry inhibition by warm temperatures in wheat under field chambers. *Physiologia Plantarum* 137, 86-100.

Leakey ADB, Bernacchi CJ, Ort DR, Long SP (2006). Long-term growth of soybean at elevated CO₂ does not cause acclimation of stomatal conductance under fully open-air conditions. *Plant Cell and Environment* 29, 1794-1800.

Miglietta F, Giuntoli A and Bindi M (1996). The effect of free air carbon dioxide enrichment (FACE) and soil nitrogen availability on the photosynthetic capacity of wheat. *Photosynthesis Research* 47, 281-290.

Mollah MR, Norton RM and Huzzey J (2009). Australian grains free air carbon dioxide enrichment (AGFACE) facility: design and performance. *Crop & Pasture Science* 60, 697 – 707.

Mpelasoka F, Hennessy K, Jones R and Bates B (2008). Comparison of suitable drought indices for climate change impacts assessment over Australia towards resource management. *International Journal of Climatology* 28, 1283-1292.

Seneweera S, Ghannoum O and Conroy JP (2001). Root and shoot factors contribute to the effect of drought on photosynthesis and growth of the C₄ grass *Panicum coloratum* at elevated CO₂ partial pressures. *Australian Journal of Plant Physiology* 28, 451-460.

Sicher RC and Bunce JA (1997). Relationship of photosynthetic acclimation to changes of Rubisco activity in field-grown winter wheat and barley during growth in elevated carbon dioxide. *Photosynthesis Research* 52, 27-38.

Zivcak M, Repkova J, Olsovska K and Brestic M (2009). Osmotic adjustment in winter wheat varieties and its importance as a mechanism of drought tolerance. *Cereal Research Communications* 37, 569-572.